



Bioorganic & Medicinal Chemistry Letters 17 (2007) 5634-5637

Bioorganic & Medicinal Chemistry Letters

Synthesis of 2,3-diaryl-1,3-thiazolidine-4-one derivatives as selective cyclooxygenase (COX-2) inhibitors

Afshin Zarghi, a,* Leila Najafnia, Bahram Daraee, Drkideh G. Dadrass and Mehdi Hedayati

^aDepartment of Pharmaceutical Chemistry, School of Pharmacy, Shaheed Beheshti University of Medical Sciences, Tehran, Iran

^bPharmaceutical Research Center, School of Pharmacy, Shaheed Beheshti University of Medical Sciences, Tehran, Iran

^cSchool of Medicine, Azad University, Shaheed Beheshti University of Medical Sciences, Tehran, Iran

^dSchool of Medicine, Endocrine Research Center, Shaheed Beheshti University of Medical Sciences, Tehran, Iran

Received 16 June 2007; revised 14 July 2007; accepted 23 July 2007 Available online 22 August 2007

Abstract—A group of 2,3-diaryl-1,3-thiazolidine-4-ones, possessing a methylsulfonyl pharmacophore, were synthesized and their biological activities were evaluated for cyclooxygenase-2 (COX-2) inhibitory activity.

© 2007 Elsevier Ltd. All rights reserved.

The use of nonsteroidal anti-inflammatory drugs (NSA-IDs) for the treatment of inflammation and pain is often accompanied by adverse gastrointestinal and renal side effects. Their anti-inflammatory activity results from inhibition of cyclooxygenases (COXs), which catalyzes the bioconversion of arachidonic acid to prostaglandins. However, inhibition of COXs may lead to undesirable side effects. Nowadays, it is well established that there are at least two COX isozymes, COX-1 and COX-2.1 The constitutive COX-1 isozyme is produced in a variety of tissues and appears to be important to the maintenance of physiological functions such as gastroprotection and vascular homeostasis.² Alternatively, the COX-2 isozyme is induced by mitogenic and proinflammatory stimuli linking its involvement to inflammatory processes.³ Thus, selective inhibition of COX-2 over COX-1 is useful for the treatment of inflammation and inflammation-associated disorders with reduced gastrointestinal toxicities when compared with NSAIDs. In addition to role of COX-2 in rheumatoid arthritis and osteoarthritis, it is also implicated in colon cancer and angiogenesis. ^{4,5} Recent studies have shown that the progression of Alzheimer's disease is reduced among some users of NSAIDs. Chronic treatment with selective COX-2 inhibitors may therefore slow the progress of

Alzheimer's disease without causing gastrointestinal damage.⁶ Diarylheterocycles, and other central ring pharmacophore templates, have been extensively studied as selective COX-2 inhibitors. All these tricyclic molecules possess 1,2 diaryl substitution on a central hetero- or carbocyclic ring system (see structures 1–5 in Chart 1).7-12 Recently, we reported several investigations describing the design, synthesis, and COX inhibitory activities of a novel class of compounds possessing an acyclic 1,3-diarylprop-2-en-1-one structural template. ^{13,14} For example, the acyclic (*E*) 1,3diphenylprop-2-en-1-ones possessing a 4-methylsulfonyl or 4-azido COX-2 pharmacophore group at the C-1 phenyl ring (see structure 6) exhibited high selective COX-2 inhibition. As part of our ongoing program to design new types of tricyclic selective COX-2 inhibitors, we now report the design, synthesis, cyclooxygenase inhibitory, and some molecular modeling studies of a group of 2,3-diaryl-1,3-thiazolidine-4-ones possessing a COX-2 SO₂Me pharmacophore at the *para*-position of C-2 phenyl ring in conjunction with a substituent (H, F, Me, and OMe) at the para-position of the N-3 phenyl ring.

The target 2,3-diaryl-1,3-thiazolidine-4-one derivatives were synthesized via the route outlined in Scheme 1. Accordingly, an appropriate aromatic amine (1) was treated with 4-methylthiobenzaldehyde (2) and thioglycolic acid (3) in dry toluene under reflux to give 2-(4-methylthiophenyl)-3-(4-substitutedphenyl)-1,3-thiazolidine-4-one (4, 20–45%) (4, 20–45%)¹⁸. Oxidation of 4 using 30%

Keywords: 1,3-Thiazolidine-4-ones; COX-2 inhibition; SAR.

* Corresponding author. Tel.: +98 21 88773521; fax: +98 21 88795008; e-mail: azarghi@yahoo.com

$$F_3C$$
 N
 Me
 SO_2NH_2
 SO_2NH_2

Chart 1. Representative examples of selective COX-2 inhibitors.

R NH₂ + OHC SMe + HS COOH Toluene

1 2 3

R SO₂ Me

4a,
$$X = H$$
4b, $X = F$
4c, $X = Me$
4d, $X = OMe$

5a, $X = H$
5b, $X = F$
5c, $X = Me$
5d, $X = OMe$
5d, $X = OMe$

Scheme 1. Reagents and conditions: (a) Toluene, reflux, 48 h; (b) $\rm H_2O_2$ 30%, $\rm WO_3$, 25 °C, 12 h.

 H_2O_2 in hydromethanol in the presence of a trace amount of WO_3 afforded the 2-(4-methylsulfonylphenyl)-3-(4-substitutedphenyl)-1,3-thiazolidine-4-one (5, 60-80%). ^{15,19}

It is well established for the diarylheterocyclic class of COX-2 inhibitors, that a *para*-methylsulfone substituent on one of the phenyl rings is a requirement for good COX-2 potency and selectivity. Accordingly, a group of 2,3-diaryl-1,3-thiazolidine-4-one analogues having different substituents at *para*-position of the N-3 phenyl ring (5a-d) were prepared to investigate the effect of different substituents on COX-2 selectivity and potency. The ability of the 1,3-thiazolidine-4-ones 5a-d to inhibit the COX-1 and COX-2 isozymes was determined using chemiluminescent enzyme assays (see enzyme inhibition data in Table 1). In vitro COX-1/COX-2 inhibition studies showed that all compounds 5a-d were selective inhibitors of the COX-2 isozyme with IC₅₀ values in the

Table 1. In vitro COX-1 and COX-2 enzyme inhibition data

Compound	R	$IC_{50}^{a} (\mu M)$		COX-2 SI ^b
		COX-1	COX-2	
5a	Н	96.1	0.20	480.5
5b	F	>100	0.12	>833
5c	Me	53.9	0.15	359.3
5d	OMe	>100	0.16	>625
Celecoxib		24.3	0.06	> 403

^a Values are mean values of two determinations acquired using an ovine COX-1/COX-2 assay kit, where the deviation from the mean is <10% of the mean value.</p>

highly potent 0.12–0.20 μM range, and COX-2 selectivity indexes (SI) in the >359 to >833 range. In addition, compounds 5b and 5d did not inhibit COX-1 at a concentration of 100 μ M (IC₅₀ values > 100 μ M). However, compounds 5a and 5c showed less selectivity for COX-2 isozyme compared with compounds 5a and 5c. According to these results, 3-(4-fluorophenyl)-2-(4-methylsulfonylphenyl)-1,3-thiazolidine-4-one 5b was the most potent (IC₅₀ = $0.12 \mu M$), and selective (SI > 833), COX-2 inhibitor among the synthesized compounds. It was also more selective COX-2 inhibitor than the parent reference compound celecoxib (SI > 403). These data suggest that the compound 5b should inhibit the synthesis of inflammatory prostaglandins via the cyclooxygenase pathway at sites of inflammation and be devoid of ulcerogenicity due to the absence of COX-1 inhibition.

The orientation of the highly potent and selective COX-2 inhibitor, 3-(4-fluorophenyl)-2-(4-methylsulfonylphenyl)-1,3-thiazolidine-4-one **5b** in the COX-2 active site,

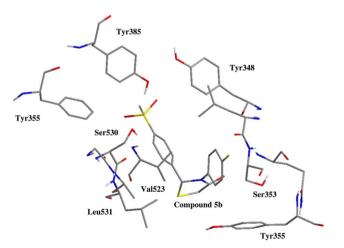


Figure 1. Compound **5b** 3-(4-fluorophenyl)-(2-(4-methylthiophenyl)-1,3-thiazolidine-4-one docked in the active site of murine COX-2 isozyme.

^b In vitro COX-2 selectivity index (COX-1 IC₅₀/COX-2 IC₅₀).

was examined by a docking experiment (Fig. 1). 16,17 This molecular modeling shows that it binds in the primary binding site such that the C-2 para-SO₂Me substituent inserts into the 2° pocket present in COX-2. One of the O-atoms of p-SO₂Me forms a hydrogen bonding interaction with hydroxyl group (OH) of Tyr³⁸⁵ (distance < 2) whereas the other O-atom is close to OH of Ser⁵³⁰ (distance = 2.7). The C=O of the central thiazolidine-4-one forms hydrogen bond (distance = 4.4) with the OH of Ser³⁵³. It was interesting to note that, the para-fluoro substituent of N-3 phenyl ring was forming a hydrogen bond with amide hydrogen (NH) of Ser³⁵³ (distance = 4.4). These observations together with experimental results provide a good explanation for the potent and selective inhibitory activity of **5b**.

Acknowledgement

We are grateful to the research deputy of Shaheed Beheshti University of Medical Sciences for financial support of this research.

References and notes

- (a) Fu, J. Y.; Masferrer, J. L.; Seibert, K.; Raz, A.; Needleman, P. J. Biol. Chem. 1990, 265, 16737; (b) Xie, W. L.; Chipman, J. G.; Robertson, D. L.; Erikson, R. L.; Simmons, D. L. Proc. Natl. Acad. Sci. U.S.A. 1991, 88, 2692.
- 2. Smith, W. L.; DeWitt, D. L. Adv. Immunol. 1996, 62, 167.
- 3. Herschman, H. R. Biochem. Biophys. Acta 1996, 1299, 125.
- Kawamori, T.; Rao, C. V.; Seibert, K.; Reddy, B. S. Cancer Res. 1998, 58, 409.
- 5. Katori, M.; Majima, M. Inflamm. Res. 2000, 295, 802.
- 6. Vane, J. R.; Botting, R. M. Inflamm. Res. 1998, 47, S78.
- Penning, T. D.; Tally, J. J.; Bertenshaw, S. R.; Carter, J. S.; Collins, P. W.; Docter, S.; Graneto, M. J.; Lee, L. F.; Malecha, J. W.; Miyashiro, J. M.; Rogers, R. S.; Rogier, D. J.; Yu, S. S.; Anderson, G. D.; Burton, E. G.; Cogburn, J. N.; Gregory, S. A.; Koboldt, C. M.; Perkins, W. E.; Seibert, K.; Veenhuizen, A. W.; Zhang, Y. Y.; Isakson, P. C. J. Med. Chem. 1997, 40, 1347.
- 8. Riendeau, D.; Percival, M. D.; Brideau, C.; Dube, C. S.; Ethier, D.; Falgueyret, J. P.; Friesen, R. W.; Gordon, R.; Greig, G.; Guay, J.; Girard, Y.; Prasit, P.; Zamboni, R.; Rodger, I. W.; Gresser, M.; Ford-Hutchinson, A. W.; Young, R. N.; Chan, C. C. J. Pharmacol. Exp. Ther. 2002, 296, 558.
- Prasit, P.; Wang, Z.; Brideau, C.; Chan, C.-C.; Charlson, S.; Cromlish, W.; Ethier, D.; Evans, J. F.; Ford-Hutchinson, A. W.; Gauthier, J. Y.; Gordon, R.; Guay, J.; Gresser, M.; Kargman, S.; Kennedy, B.; Leblanc, Y.; Leger, S.; Mancini, J.; O_Neil, G. P.; Quellet, M.; Percival, M. D.; Perrier, H.; Riendeau, D.; Rodger, I.; Tagari, P.; Therien, M.; Vikers, P.; Wong, E.; Xu, L.-J.; Young, R. N.; Zamboni, R.; Boyce, S.; Rupniak, N.; Forrest, M.; Visco, D.; Patrick, D. Bioorg. Med. Chem. Lett. 1999, 9, 1773.
- Friesen, R. W.; Dube, D.; Fortin, R.; Frenette, R.; Prescott, S.; Cromlish, W.; Greig, G. M.; Kargman, S.; Wong, E.; Chan, C. C.; Gordon, R.; Xu, L. J. Bioorg. Med. Chem. Lett. 1996, 6, 2677.
- 11. Talley, J. J.; Brown, D. L.; Carter, J. S.; Graneto, M. J.; Koboldt, C. M.; Masferrer, J. L.; Perkins, W. E.; Rogers,

- R. S.; Shaffer, A. F.; Zhang, Y. Y.; Zweifel, B. S.; Seibert, K. J. Med. Chem. 2000, 43, 775.
- Zarghi, A.; Rao, P. N. P.; Knaus, E. E. J. Pharm. Pharm. Sci. 2007, 10, 129.
- Zarghi, A.; Arfaee, S.; Rao, P. N. P.; Knaus, E. E. *Bioorg. Med. Chem.* 2006, 14, 2600.
- Zarghi, A.; Zebardast, T.; Hakimion, F.; Shirazi, H. F.;
 Rao, P. N. P.; Knaus, E. E. *Bioorg. Med. Chem.* 2006, 14, 7044
- Cho, I. H.; Lim, J. W.; Park, S. W.; Noh, J. Y.; Ryu, H. C.; Kim, J. H.; Chae, M. Y.; Park, H. J.; Jung, S. H.; Yeon, K. J.; Jin, H. T.; Lee, Y. P. U.S. Patent WO/2003/097620, 2003.
- 16. Docking studies were performed using Autodock software Version 3.0. The coordinates of the X-ray crystal structure of the selective COX-2 inhibitor SC-558 bound to the murine COX-2 enzyme was obtained from the RCSB Protein Data Bank (1c×2) and hydrogens were added. The ligand molecules were constructed using the Builder module and were energy minimized for 1000 iterations reaching a convergence of 0.01 kcal/mol Å. The energy minimized ligands were superimposed on SC-558 in the PDB file 1c×2 after which SC-558 was deleted. The purpose of docking is to search for favorable binding configuration between the small flexible ligands and the rigid protein. Protein residues with atoms greater than 7.5 Å from the docking box were removed for efficiency. Searching is conducted within a specified 3D docking box using annealing based on the Monte Carlo method and MMFF94 molecular mechanics force field for 8000 iterations. These docked structures were very similar to the minimized structures obtained initially. The quality of the docked structures was evaluated by measuring the intermolecular energy of the ligand-enzyme assembly.
- 17. Kurumbail, R. G.; Stevens, A. M.; Gierse, J. K.; McDonald, J. J.; Stegeman, R. A.; Pak, J. Y.; Gildehaus, D.; Miyashiro, J. M.; Penning, T. D.; Seibert, K.; Isakson, P. C.; Stallings, W. C. *Nature* **1996**, *384*, 644.
- 18. Analytical data for **4a**. Yield, 45%; white crystalline powder; mp 105–106 °C; IR (KBr): v cm⁻¹ 1680 (C=O); MS: m/z (%): 301 (M⁺, 60), 226(55), 177(75), 134(100), 104(45), 91(30), 77(40); 1 H NMR (CDCl₃): δ ppm 2.48 (S, 3H, SCH₃), 3.90 (d, 1H, CH₂, J = 15.8 Hz), 4.04 (d, 1H, CH₂, J = 15.8 Hz), 6.10 (S, 1H, CH), 7.18 (d, 2H, 4-methylthio phenyl H₃ and H₅, J = 8.4 Hz), 7.19–7.23 (m, 3H, phenyl H₃—H₅), 7.25 (d, 2H, 4-methylthio phenyl H₂ and H₆, J = 8.4 Hz), 7.33 (dd, 2H, phenyl H₂ and H₆, J = 7.8 Hz); Anal. C₁₆H₁₅NOS₂ (C, H, and N).
 - Compound **4b**. Yield, 35%; white crystalline powder; mp 97 °C; IR (KBr): v cm⁻¹ 1670 (C=O); MS: m/z (%): 319 (M⁺, 30), 243(40), 228(20), 177(45), 134(100), 94(70), 77(40); Anal. $C_{16}H_{14}FNOS_2$ (C, H, and N).
 - Compound **4c**. Yield, 20%; white crystalline powder; mp 101-102 °C; IR (KBr): $v \text{ cm}^{-1}$ 1680 (C=O); MS: m/z (%): 315 (M⁺, 60), 240(70), 178(100), 135(90), 91(90), 77(30); Anal. C₁₇H₁₇NOS₂ (C, H, and N).
 - Compound **4d**. Yield, 35%; white crystalline powder; mp 103-104 °C; IR (KBr): v cm⁻¹ 1670 (C=O); MS: m/z (%): 331 (M⁺, 25), 257(45), 242(50), 178(45), 135(100), 77(30); Anal. $C_{17}H_{17}NOS_2$ (C, H, and N). Satisfactory analysis for C, H, N was obtained for all the compounds within $\pm 0.4\%$ of the theoretical values.
- 19. Analytical data for **5a**. Yield, 60%; white crystalline powder; mp 238–239 °C; IR (KBr): $v \text{ cm}^{-1}$ 1670 (C=O), 1300, 1120 (SO₂); MS: m/z (%): 333 (M⁺, 15), 259(45), 179(30), 123(70), 77(100); ¹H NMR (CDCl₃): δ ppm 3.23 (S, 3H, SO₂CH₃), 3.92 (d, 1H, CH₂, J = 16.5 Hz), 4.08 (d, 1H, CH₂, J = 16.5 Hz), 6.71 (S, 1H, CH), 7.25–7.50 (m, 5H, phenyl), 7.82 (d, 2H, 4-methylsulfonyl phenyl H₂ and

 H_6 , J = 8.4 Hz), 8.01 (d, 2H, 4-methylsulfonyl phenyl H_3 and H_5 , J = 8.4 Hz); Anal. $C_{16}H_{15}NO_3S_2$ (C, H, and N). Compound 5b. Yield, 65%; white crystalline powder; mp 234 °C; IR (KBr): ν cm⁻¹ 1670 (C=O), 1310, 1120 (SO₂); MS: m/z (%): 351 (M⁺, 5), 277(100), 197(60), 122(60), 95 (90), 77(50); ¹H NMR (CDCl₃): δ ppm 3.24 (S, 3H, SO_2CH_3), 3.74 (d, 1H, CH_2 , J = 17.0 Hz), 4.41 (d, 1H, CH_2 , J = 17.0 Hz), 6.66 (S, 1H, CH), 7.27 (t, 2H, 4fluorophenyl H₃ and H₅, J = 8.8 Hz), 7.53 (dd, 2H, 4fluorophenyl H_2 and H_6 , $J_{HH} = 8.6 \text{ Hz}$, $J_{HF} = 4.9 \text{ Hz}$), 7.81 (d, 2H, 4-methylsulfonyl phenyl H₂ and H₆, J = 8.3 Hz), 7.99 (d, 2H, 4-methylsulfonyl phenyl H₃ and H_5 , J = 8.3 Hz); Anal. $C_{16}H_{14}FNO_3S_2$ (C, H, and N). Compound 5c. Yield, 75%; white crystalline powder; mp 236–237 °C; IR (KBr): υ cm⁻¹ 1680 (C=O), 1300, 1110 (SO₂); MS: m/z (%): 347 (M⁺, 5), 273(100), 193(60), 118(60), 91 (100), 77(50); ¹H NMR (CDCl₃): δ ppm 2.26 (S, 3H, CH₃), 3.23 (S, 3H, SO₂CH₃), 3.71 (d, 1H, CH₂, J = 17.0 Hz), 4.39 (d, 1H, CH₂, J = 17.0 Hz), 6.67 (S, 1H, CH), 7.21 (d, 2H, 4-methylphenyl H_3 and H_5 , J = 8.4 Hz), 7.38 (d, 2H, 4-methylphenyl H_2 and H_6 , J = 8.4 Hz), 7.80 (d, 2H, 4-methylsulfonyl phenyl H_2 and H_6 , J = 8.3 Hz), 7.98 (d, 2H, 4-methylsulfonyl phenyl H₃ and H₅, J = 8.3 Hz); Anal. $C_{17}H_{17}NO_3S_2$ (C, H, and N). Compound 5d. Yield, 80%; white crystalline powder; mp 205–206.5 °C; IR (KBr): υ cm⁻¹ 1670 (C=O), 1300, 1100 (SO₂); MS: m/z (%): 363 (M⁺, 5), 289(100), 274(75), 195(40), 91 (90), 77(30); ¹H NMR (CDCl₃): δ ppm 3.23 (S, 3H, SO_2CH_3), 3.70 (d, 1H, CH_2 , J = 17.2 Hz), 3.75 (S, 3H, OCH₃), 4.38 (d, 1H, CH₂, J = 17.2 Hz), 6.65 (S, 1H, CH), 6.96 (d, 2H, 4-methoxyphenyl H₃ and H₅, J = 8.9 Hz), 7.37 (d, 2H, 4-methoxyphenyl H₂ and H₆, J = 8.9 Hz), 7.81 (d, 2H, 4-methylsulfonyl phenyl H₂ and H_6 , J = 8.3 Hz), 7.98 (d, 2H, 4-methylsulfonyl phenyl H₃ and H₅, J = 8.3 Hz); Anal. C₁₇H₁₇NO₄S₂ (C, H, and N). Satisfactory analysis for C, H, N was obtained for all the compounds within ±0.4% of the theoretical values.